Total carbohydrate content and changes therein in seeds of Schleichera oleosa (kusum) due to biodeterioration by pathogenic fungi during storage

A.K. SRIVASTAVA*, SANYUKTA KUMAR** AND GOVIND K. PANDEY

Department of Botany, St. Columba's PG College, Hazaribag, *Department of Botany, St. Xavier's College, Ranchi and **Department of Biotechnology, St. Xavier's College, Ranchi

Received: 25.02.2010

Accepted: 24.06.2011

Published: 24.10.2011

Seeds of one of the non-edible oilseed, kusum (*Schleichera oleosa*, Sapindaceae) are of great significant in Jharkhand area and its oil is used in burning lamps, varnishing, massage and medicine. Its oil-cake is a good manure. The unsanitary and humid condition makes it proned to a faster biodeterioration due to several fungal inhabitants during storage. The colonising fungi in the seeds used up some of the glucose, sugars and starch as carbon sources for meeting energy requirements in their prolonged association with the seeds. In the span of one year the glucose level dropped by as much as 50.2 per cent of its initial value. Similarly, totall sugar also registered a loss of 40.4 per cent. The starch in the stored seeds reduced steadily from 2.55 per cent of dry seed weight to 1.25 per cent due to the association of the mycodwellers.

Key words: Schleichera_oleosa, oilseed, colonising fungi, carbon sources, mycodwellers

INTRODUCTION

Seed pathology is an up coming area of plant pathology and has attained a great magnitude. Seeds of a non edible oil, *Schleichera oleosa*. Sapindaceae (kusum) are of great use in burning lamps, varnishing, massage and medicine. Its oilcake is good manure. In the Jharkhand area, kusum seeds are collected just after onset of rainy season i.e. by the end of month of May. Conventionally seeds are stored in gunny bags in the village houses and godowns. In either case the condition is unsanitary and humid and is conducive for fungal growth leading to good scale destruction.

There are many reports indicating that fungi are associated with seed surface as well as internal mycoflora. It is worthwhile to seek the extent of biodeterioration occurring in the seeds in terms of carbohydrates in different forms viz. glucose, total sugars and starch.

MATERIALS AND METHODS

Kusum seeds were obtained from the godown of Ranchi forest department for the preparation of oil and oilcake on regular interval of one month. The work spanned from June 2001, one harvest season to May 2002, just before the other harvest season. Glucose content was estimated by titration method using Fehling solutions. Total sugars and starch were estimated by colorimetric method using anthrone as a reagent (Dubois *et al.*, 1951) using a Colorimeter of Systronics (India) make. Estimations were done in triplicate and standard deviation and standard errors were calculated.

RESULTS AND DISCUSSION

The effect of continued fungal activity on the seeds manifested in the lowered glucose, total soluble sugars and starch contents just like oils (Srivastava and Pandey 2000). The glucose level dropped by as much as 50.2 per cent in the span of one year of its initial value. Similarly, total sugar also registered a loss of 40.4 per cent. The starch in the stored seeds reduced steadily from 2.55 per cent of dry sed weight to 1.25 per cent due to the association of the mycodwellers.

Changes in glucose content with pathogenic association have been reported. Loss in amount of glucose in fruits have been reported for tomato-Drechslera australiense (Kapoor and Tandon 1970); tomato - Alternaria solani (Mehta et. al.,

Table 1 : Amount of glucose present in 1 g of Kusum seed powder samples through one year of biodeterioration due to fungal attack, June '01- May '02.

Month	Wt.(mg)	Percentage	Change (mg)
Jun '01	16.75	1.67	
Jul '01	16.25	1.62	-0.05
Aug '01	16.25	1.62	-0.05
Sept '01	15.75	1.57	-0.1
Oct '01	14.9	1.49	-0.17
Nov '01	14.3	1.43	-0.24
Dec '01	13.1	1.31	-0.36
Jan '02	13.0	1.3	-0.37
Feb '02	11.9	1.19	-0.48
Mar '02	10.3	1.03	-0.64
Apr '02	9.5	0.95	-0.72
May '02	8.35	0.83	-0.84
S.D. 0.284	S.E. 0.081		

Final per cent change in glucose content : [(1.675-0.835)/1.675] 100 = 50.2%

Table 2: Amount of sugars present in 1 g of kusum seed powder samples through one year of biodeterioration due to fungal attack, June '01 - May '02

Month	Wt.(mg)	Percentage	Change (mg)
Jun '01	47	4.7	
Jul '01	47	4.7	- 0
Aug '01	45	4.5	- 0.2
Sept '01	44	4.4	- 0.3
Oct '01	41	4.1	- 0.6
Nov '01	40	4.0	- 0.7
Dec '01	37	3.7	- 1.0
Jan '02	35	3.5	- 1.2
Feb '02	34	3.4	- 1.3
Mar '02	32	3.2	- 1.5
Apr '02	30	3.0	- 1.7
May '02	28	2.8	- 1.9
S.D. 0.663	S.E. 0.191		

Final per cent change in sugar content: [(47-28)/47] 100 = 40.425%

1975); and banana-Gloeosporium musarum, (Wang 1960).

Changes in total sugar content in the host tissue due to pathogenic actions have been reported by Craig and Hooker (1961); Dhanvantari (1967), Dayal and Joshi; (1968) and Padmanabhan, et al. (1988) on different host pathogen systems.

Similarly the depletion in the starch content is commensurate with the activation of starch degrading enzymes (Schipper and Mirocha,

1977). The drop is due to the sugar being used by the fungal pathogens as respiratory substrates (Baker, 1965; Wu, 1973). In pigeon pea seeds infested with Aspergillus flavus, Sinha and Prasad (1977) have found a depletion of starch. Likewise Bilgrami et al. (1979) have recorded a considerable decrease in the amount of strach in paddy seeds during 60 and 90 days of fungal infestation by an aflatoxin producing strain of Aspergillus parasiticus. Sinha et al., (1981) have found a considerable reduction in the starch content of Cajanus cajanseeds infested with Aspergillus flavus and A. niger. However, these seeds, when infested with Alternaria alternata and Curvularia lunata, have shown a moderate reduction in the starch content. On the contrary, Cajanus-seeds infested with Fusarium moniliforme and Drechslera hawaiiensis have exhibited a minimum level reduction in starch contents. Later on, Singh and Sinha (1985) have confirmed that infestation of Cajanus seeds by Aspergillus parasiticus caused a considerable decline in their starch contents. Prasad (1989) has reported a loss of starch in fungi infested seeds of Coriandrum indicum and the maximum loss in starch has been due to Aspergillus flavus followed by Curvularia lunata.

All these reports are concurrent to our findings. It is thus appearnt that one year of seed infestation of *Scheleichera oleosa* predominantly and jointly by *Aspergillus fumigatus*, *A. flavus*, *A. niger*, *Fusarium solanii*, *Paecilomyces variotii* and *Mucor*

Table 3: Amount of starch present in 1 g of kusum seed powder samples through one year of biodeterioration due to fungal attack, June '2001 - May '02

Month	Wt.(mg)	Percentage	Change (mg)
Jun '01	25.5	2.55	
Jul '01	24.0	2.4	- 0.15
Aug '01	22.3	2.23	- 0.22
Sept '01	22.0	2.2	- 0.25
Oct '01	21.6	2.16	- 0.39
Nov '01	19.9	1.99	- 0.56
Dec '01	17.9	1.79	- 0.76
Jan '02	17.1	1.71	- 0.84
Feb '02	16.3	1.63	- 0.92
Mar '02	14.8	1.48	- 1.07
Apr '02	12.9	1.29	- 1.26
May '02	12.5	1.25	- 1.30
S.D. 0.430	S.E. 0.124		

Final per cent change in starches content : [(25.5-12.5)/25.5] 100 = 50.98%

sp. causes biodeterioration of seeds manifested in the diminishing of carbohydrate contents.

REFERENCES

- Baker, J. 1965. Study in the respiratory and carbohydrate metabolism of plant tissues. XVIII. The effect of oxygen on starch formation and dissolution in potatoes. New Phytol. 64: 201-209.
- Bilgrami, K.S., Jamuluddin., Sinha, R.K. and Prasad, T. 1979. Changes in seed contents of paddy (Oryza sativa L) due to fungal flora. Phytopath J. 96: 9-14.
- Chattopadhyay, N.C., and Nandi, B. 1978. Changes in nitrogen content in malformed inflorescence of mango caused by Fusarium moniliforme Sheld. var subglutinans Wr. Et. Rg Giornale Bot. Ital. 112: 343-346.
- Craig, J. and Hooker, A.L. 1961. Relation of sugar trends and pith density to Diplodia stalk rot in dent corn. *Phytopathology* 51: 376-382
- Dayal, R. and Joshi, M.M. 1968. Post infection changes in the sugar content of leaf spot infected barley. *Indian Phytopath.* 21: 221-222
- Dubois, M., Gilles, K., Hamilton, J.K., Rebers, P.A. and Smith, F., 1951 A colorimetric method for the estimation of sugar. *Nature* 168: 167.
- Dhanvantri, BN. 1967. The leaf scorch disease of Strawberry, Diplocarpon earlianum and the nature of resistance to it. *Can. J. Bot.* **45**: 1525-1543
- Kapoor, IJ, and Tandon, R.N.1970. Post infection changes in sugar content of tomato fruits caused by *Dreschlera australiense*. *Indian Phytopath.* 23: 133-135.

- Mehta, P., Vyas, K.M. and Saksena, S.P. 1975. Metabolic changes during pathogenesis of fruit rot disease of tomato. *Indian Phytopath.* **28**: 253-255.
- Padmanabhan, P., Alexander, KC. and Shanmugam, N. 1988. Some metabolic changes induced in sugarcane by *Ustilago scitaminea*. *Indian Phytopath*. **41**: 229-232.
- Prasad, M, 1989. Plant-bacteria interaction- Its dynamics and dimensions. In *Plant Microbe Interactions*. Ed: K.S. Bilgrami. Narendra Publishing House, Delhi: 143-161.
- Schipper, AL and Mirocha, C.J. 1977. Mechanism of starch hydrolysis in bean leaves during infection by bean rust fungus. *Phytopathology* **58**: 1066.
- Singh, Premlata and Sinha, K.K. 1985. Changes in the seed content of arhar infected with Aspergillus parasiticus. Indian Phytopathol. 38: 560.
- Sinha, M.K. and Prasad, T., 1977. Deterioration of arhar seeds by Aspergillus flavus. Indian Phytopathol. 30: 70-72.
- SInha, M.K., Singh, K.K. and Prasad, T. 1981. Changes in starch contents of Arhar seeds due to fungi. *Indian Phytopathol.* 34: 269-271.
- Srivastava, A.K. and Pandey, G.K. 2000. Chemical changes in properties of kusum (Schleichera oleosa) oil during its seed infestation by fungi. J. Mycopathol. Res. 38(1): 29-32.
- Wang, M.C. 1960. Physiological studies on *Gloeosporum musarum* Cook. At Mass, the causal organism of banana anthracnose. Changes in the carbohydrate composition of banana pulp with reference to the adaptive secretion of *amylase*. *Bot. Bull. Acad. Sin. N.S.* 1: 59-75.
- WU, LC. 1973. Changes in some enzymes of moong bean seeds germinated on mycelia macerate of *Rhizoctonia solani*. *Physiol Plant Pathol*. 3: 19-27.